## Broad glutamine pathway inhibition by DRP-104 results in anti-tumor activity in hypermetabolic lung tumors resistant to PD-1 or osimertinib therapy. Morgan Brady<sup>1</sup>, Milica Momcilovic<sup>1</sup>, Chiara Montemurro<sup>1</sup>, Yumi Yokoyama<sup>7</sup>, Heather Christofk<sup>3,4,5</sup>, Katerina Politi<sup>6</sup>, Margaret Dugan<sup>7</sup>, Aaron Lisberg<sup>2,5</sup>, Robert Wild<sup>7</sup>

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## **Abstract:**

Recent breakthroughs with checkpoint inhibitors (CPI) or the third generation tyrosine kinase inhibitor (TKI) osimertinib (OSI) for patients with specific mutations to the epidermal growth factor receptor (EGFR) gene are now able to induce durable responses in non-small cell lung cancer (NSCLC) patients<sup>1,2</sup>. However, the majority of patients who receive CPI or OSI will be either non-responsive to treatment or develop therapy resistance. We sought to identify alternative treatment strategies to overcome therapy resistance by targeting metabolism in aggressive tumors. The fast rate of growth in therapy-resistant hyper-metabolic tumors (HMTs) makes them dependent on nutrients to sustain anabolic growth. To demonstrate this dependency, we coupled PET imaging using <sup>18</sup>F-FDG and <sup>11</sup>C-Glutamine (<sup>11</sup>C-GLN) with in vivo metabolomics and identified a conserved metabolic signature in which lung squamous cell carcinomas (LUSC) and EGFR mutant lung adenocarcinomas (LUAD) were marked by dependence upon both glucose and glutamine<sup>3,4</sup>. Importantly, this metabolic signature is predictive of either response or resistance to targeted therapies that inhibit glucose and glutamine metabolism that may be exploited in a clinical setting.

We previously reported that combined targeting of glucose and glutamine metabolism with mTOR kinase inhibitor TAK228 or TKI in combination with the selective glutaminase (GLS) inhibitor CB-839 was required to significantly suppress glucose and glutamine metabolism in both LUSC and EGFR mutant LUADs resulting in metabolic crisis and tumor cell death<sup>3,5</sup>. Here, we tested the novel broad acting glutamine antagonist DRP-104 (sirpiglenastat) in mouse models of lung cancer as a single agent in OSI-resistant EGFR mutant LUAD or in combination with anti-PD-1 in CPI-resistant LUSC.

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Figure 1. Structure, function, and targets of DRP-104 (sirpiglenastat). (A) DRP-104 is a prodrug of the broad acting glutamine antagonist DON (6-Diazo-5-oxo-L-norleucine). DRP-104 is inactive in its prodrug form with high plasma and GI tissue stability. DRP-104 is preferentially distributed in tumors where it is bio-transformed and activated to the active moiety DON. (B) Multiple glutamine dependent pathways are inhibited by DON (Lemberg KM, et al., Mol Cancer Ther 2018) (C) Model of MOA of DRP-104.



Figure 2. Glucose and glutamine dependencies in hypermetabolic tumors (HMTs). Proposed mechanism of DRP-104, directly inhibiting glutaminolysis and indirectly impacting glycolysis (Leone RD, et al., Science 2019).

Figure 4. Evaluation of DRP-104 in patient derived xenografts of LUSC and OSI-resistant EGFR-mutant LUAD. (A and B) LUSC PDX007 and PDX005 were treated with vehicle, 1mg/kg DRP-104, or 3mg/kg DRP-104 for 9 days (PDX007) or 13 days (PDX005). Top images are representative images of tumors stained for antibodies against GLUT1 and SLC1A5<sup>3</sup>. Graphs show tumor growth curve. Tumor volumes were measured three times per week with calipers. For all experiments mice received daily treatments either vehicle or DRP-104 (1 or 3mg/kg/s.c./q.d.) 5 days ON, 2 days OFF. (C) EGFR-mutant LUAD PDX-YLR-086 is resistant to OSI treatment and responsive to treatment with DRP-104. PDX-YLR086 was treated with vehicle, 1.4mg/kg DRP-104, 3mg/kg DRP-104, 25mg/kg OSI, or combination of 1.4mg/kg DRP-104 and 25mg/kg OSI. Graph shows tumor growth curve. Tumor volumes were measured with calipers three times per week. For all experiments mice received daily treatments either vehicle, DRP-104 (1.4 or 3mg/kg/s.c./q.d.) 5 days ON, 2 days OFF, or OSI (25mg/kg/p.o./q.d.). The data are represented as the mean +/-SEM. Statistical significance (\*p<0.05; \*\*p<0.01; \*\*\*<0.001; \*\*\*\*<0.0001) and analyzed via one-way ANOVA with post-hoc Dunnett's correction.



PDX00 PDX00 RH2 HCC **YLRO** A54 H143 H194 PDX013 PDX0

(A and B) Growth curve representing mice treated with Vehicle, DRP-104, anti-PD1, or a combination of DRP-104 and anti-PD1 for 16 days (3605 T1) or 18 days (1969b). Growth measured by caliper 3 times a week. All mice received Vehicle, DRP-104 (1mg/kg/s.c./q.d.), anti-PD1 (10mg/kg/i.p./q.4d.), or a combination of DRP-104 (1mg/kg/s.c./g.d.) and anti-PD1 (10mg/kg/i.p./g.4d.). DRP-104 was provided 5 days ON 2 days OFF. The data are represented as the mean +/- SEM. Comparisons between treatment and vehicle noted. Statistical significance (\*p<0.05; \*\*p<0.01; ns, not significant) and analyzed via one-way ANOVA with post-hoc Dunnett's correction.

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	Histology	Туре	Identified Genetic Mutations	Response to DRP-104
Human Xenograft and PDX Tumor Models				
07	LUSC	PDX	<b>KEAP1<sup>M317V, V316A, E306</sup></b> Q	Responder
05	LUSC	PDX	KEAP1 <sup>N438S, R362Q, S275A</sup> /p53 <sup>V41L, Q33K, P33R</sup> /PTEN <sup>A97S</sup>	Partial responder
2	LUSC	Xenograft	KRAS <sup>Q61H</sup> /LKB1 <sup>Q37X</sup> /KEAP1 <sup>D236H</sup> /p53 <sup>C33R</sup>	Partial responder
15	LUSC	Xenograft	LKB1 (large del)/KEAP1 <sup>G364C</sup> /p53 <sup>D259V</sup>	Partial responder
86	LUAD	PDX	EGFR <sup>L747_A750</sup> (del19)	Responder
9	LUAD	Xenograft	KRAS <sup>G12S</sup> /LKB1 <sup>Q37</sup> /KEAP1 <sup>G333C</sup>	Partial responder
37	LUAD	Xenograft	<i>LKB1</i> (p.E98-G155 del)	Partial responder
14	LUAD	Xenograft	KRAS <sup>G12C</sup> /LKB1 <sup>K62N</sup> /KEAP1 <sup>R272L</sup>	Partial responder
3-CR	LUAD	PDX	KEAP1 <sup>G417V, M317V, V316A, E306Q</sup> /PTEN <sup>C65S, S89P, A97S, S129G</sup>	Partial responder
13	LUAD	PDX	KEAP1 <sup>G417V</sup> /PTEN <sup>V38L, C65S, S129G</sup>	Non-responder
58	LUAD	Xenograft	<i>LKB1</i> (del)	Non-responder
Mouse Syngeneic Tumor Models				
T1	LUSC	Syngeneic cell line	Lkb1 <sup>lox/lox</sup> /p53 <sup>lox/lox</sup> /Pten <sup>lox/lox</sup>	Responder
)b	LUAD	Syngeneic cell line	Kras <sup>G12D</sup> /p53 <sup>lox/lox</sup>	Responder
2-B8	LUAD	Syngeneic cell line	Kras <sup>G12D</sup> /p53 <sup>lox/lox</sup> /Lkb1 <sup>lox/lox</sup>	Non-responder
1c	LUAD	Syngeneic cell line	Kras <sup>G12D</sup> /p53 <sup>lox/lox</sup> /Lkb1 <sup>lox/lox</sup>	Non-responder

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## **Conclusions:**

DRP-104 (sirpiglenastat) significantly inhibited tumor progression in the 3605 T1 (*Lkb1<sup>-/-</sup>;Pten<sup>-/-</sup>;p53<sup>-/-</sup>*) and 1969b (KRAS<sup>G12D</sup>;p53<sup>-/-</sup>) genetically engineered mouse tumor models (GEMMs) but not in the related GEMM models 2785 2-B8 (Kras<sup>G12D</sup>;p53<sup>-/-</sup>;Lkb1<sup>-/l-</sup>) and L3161c (Kras<sup>G12D</sup>;p53<sup>-/-</sup>;Lkb1<sup>-/l-</sup>). Responsive tumors had a higher KEAP1/NRF2 pathway score compared to non-responsive tumors. Interestingly, single agent anti-tumor activity of DRP-104 was extended into several xenograft and PDX tumor models with KEAP1 mutations, including models co-harboring mutations in KRAS, PTEN, p53 and/or LKB1. In addition, we showed that DRP-104 induced a significant response in an OSI-resistant PDX model of EGFR mutant LUAD. Lastly, in the DRP-104 responsive but anti-PD-1 non-responsive GEMM tumors, the combination of DRP-104 and anti-PD-1 treatment demonstrated therapeutic synergy, suggesting that broad inhibition of glutamine metabolism by DRP-104 induced metabolic remodeling of the tumor immune microenvironment and permissiveness to CPI blockade<sup>6</sup>. Overall, DRP-104 demonstrated potential to treat hypermetabolic therapy-resistant LUSC and LUAD as a single-agent therapy and in combination with immune CPI. Further clinical development of DRP-104 (sirpiglenastat) in this patient population is warranted and a first-in-human clinical trial is currently ongoing.

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